Effects of Sintering Temperature on Crystallization and Fabrication of Porous Bioactive Glass Scaffolds for Bone Regeneration

E. P. Erasmus1,2,3, O. T. Johnson1,4, I. Sigalas1,2,3 & J. Massera5

In this work the sintering ability of borosilicate (S53B50), borophosphate (P40B10) and phosphate (Sr) bioactive glasses was investigated. The glass powders were crushed and sintered in air at a heating rate of 10 °C/min for 2 hours at sintering temperatures between 480 °C–600 °C. The aim was to define the optimum sintering temperature prior to glass crystallization. The density of the samples was found to decrease when the temperature was increased up to 580 °C, probably due to the inhibition of the viscous flow of the particles during sintering thereby reducing the densification of the material. Such low porosity is not suitable in tissue engineering. To process highly porous scaffolds with porosity required for scaffold applicable to tissue engineering, the powders were further mixed with 60 vol.% and 70 vol.% of NH4(HCO3) foaming agent. Meanwhile, the density of the samples sintered with NH4(HCO3) was found to decrease with an increase in NH4(HCO3) content. This indicates an increase in porosity of the samples. The glass compositions reached an open porosity of more than 60% at the addition of 70 vol.% NH4(HCO3). In addition, SEM micrograph revealed large pores with good interconnection between the pores.

The large number of bone defects caused from a disease or trauma still remains a significant clinical challenge. At present, autografts and allografts are mainly used in the repair of defected bones. These grafts offered all the desired properties required for bone repair and regeneration, which are osteoconductivity, osteogenesis, and osteoinductivity. However, bone grafts are associated with high operating costs for harvesting the graft, limited availability, donor site morbidity and complications including infection, pain, and hematoma1,2. On the other hand, allografts are subject to cleaning and preparation processes designed to remove cells to minimize immune response. In addition, these process techniques potentially reduce the osteoinductivity, osteoconductivity, and mechanical strength of the graft3. Therefore investigation of biomaterials which could be used in the repair and regeneration of bones remains of paramount importance.

One such biomaterial is bioglass® (45S5) with composition in the SiO2-CaO-Na2O-P2O5 system discovered by Hench et al.4 Bioglass®, as well as other similar glass compositions, such as BonAlive(R) (S53P4) were found to rapidly form a chemical bond with bone through the precipitation of a hydroxyapatite layer at the glass surface. The mechanism enabling strong bone/glass interface has been reviewed in detail in the past5. However, typical silicate bioactive glasses have two major drawbacks despite their bond forming abilities. These include their poor mechanical properties (caused by their amorphous nature) and their tendency to crystallize during sintering6,7. According to Filho et al.8, the presence of crystalline phases formed during heat treatment tend to retard the formation of the hydroxyapatite layer, hence glass becomes less bioactive in simulated body fluid. Therefore, crystallization should be prevented whenever possible. This becomes an issue to overcome, as in order to obtain clinically relevant construct, the graft should be a 3D structure with large pore size (>100 µm) and porosity (>70%).
The construct should have mechanical properties similar to the tissue to be replaced and should be prepared in a reproducible manner. Typical 3D scaffold manufacturing involves a firing and sintering step which will lead, in most cases, to crystallization of the glass. Over the years new glass compositions with no or lower SiO₂ content have been developed. Phosphate and borophosphate glasses have been found to not only be promising as biomaterials, but also allows hot forming at temperature below their crystallization domain. Borosilicate glasses also demonstrated to have thermal properties enabling hot forming more readily than typical silicate bioactive glasses. Not only these glasses have enhanced hot forming domain but they were also found to promote cell adhesion and proliferation as well as differentiation into bone cells.

Locs et al. studied the preparation of porous ceramics by viscous slurry foaming using hydroxyapatite powder and ammonium hydrogen carbonate as the foaming agent for the generation of pores in the glycerol-based viscous ceramic slurry. Open and total porosity were found to increase from 25 to 69% and from 32 to 73% when NH₄(HCO₃) content was varied from 0 to 2.75 wt.%. The sintered ceramics were found to have well-connected, irregularly shaped pores. Meanwhile, Kazutaka et al. produced highly porous alumina based ceramics by incorporating polymethylmethacrylate (PMMA) microspheres with different diameters as a template and MgO or SiC powder as a sintering aid and subsequent calcinations at 1600 °C. The sintered ceramics formed spherical pores which correlated to the morphology of the PMMA microspheres. Highly porous and mechanically strong alumina-based ceramics having an open porosity of 62%, a connected space size of 1.3 μm, and a compressive strength of 147.6 MPa were fabricated by incorporating PMMA microspheres with a mean particle size of 22.6 μm and an appropriate amount of SiC.

This work aims to determine the time-temperature conditions that allow for the sintering of a borosilicate (S53B50), borophosphate (P40B10) and phosphate (Sr) with minor or no crystallization as well as the fabrication of porous scaffolds using NH₄(HCO₃).

### Results

**Thermal properties.** The three glasses under investigation were processed with the composition presented in Table 1.

<table>
<thead>
<tr>
<th>Composition (mol.%)</th>
<th>S53B50</th>
<th>P40B10</th>
<th>Sr</th>
</tr>
</thead>
<tbody>
<tr>
<td>SiO₂</td>
<td>26.93</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Na₂O</td>
<td>22.66</td>
<td>10.00</td>
<td>10.00</td>
</tr>
<tr>
<td>CaO</td>
<td>21.77</td>
<td>20.00</td>
<td>0</td>
</tr>
<tr>
<td>B₂O₃</td>
<td>26.93</td>
<td>10.00</td>
<td>0</td>
</tr>
<tr>
<td>P₂O₅</td>
<td>1.72</td>
<td>40.00</td>
<td>50.00</td>
</tr>
<tr>
<td>SrO</td>
<td>0</td>
<td>20.00</td>
<td>40.00</td>
</tr>
</tbody>
</table>

| Figure 1. | DTA thermograph of the glasses of investigation. |

The three glasses under investigation were processed with the composition presented in Table 1.

Figure 1 shows the DTA trace of the glasses S53B50, P40B10 and Sr recorded at a heating rate of 10 °C/min on a powder sample of 125–250 μm particle size. The glass transition (Tₐ) is taken at the inflection point of the endotherm (obtained by taking the first derivative of the curve) and Tₓ is defined as the onset of the crystallization peak. The working range (or hot forming domain) was defined as ΔT = Tₓ − Tₐ. In this study, the crystallization temperature (Tₓ) which is at the maximum of the exothermic peak was not measured. The quantified values are presented in Table 2.
Powder morphology of bioactive glasses. Figure 2 shows the XRD patterns of the as prepared powders prepared without $\text{NH}_4(\text{HCO}_3)$. The patterns showed that the bioactive glasses were amorphous and free of any significant crystalline phases.

Figure 3 presents the SEM micrographs of the powders. It reveals that the bioactive glass powders were composed of particles exhibiting irregular shapes and a wide size distribution, with small particles tending to attach themselves to the surfaces of the larger ones. The average particle size obtained after milling for glasses S53B50, P40B10 and Sr were 20.54 $\mu$m, 18.37 $\mu$m and 26.37 $\mu$m respectively. EDS analysis was performed and the glass composition was found to be in agreement with the nominal one (Table 1).

Similar results were observed for the bioactive glass powders prepared with $\text{NH}_4(\text{HCO}_3)$, it is important to note that the $\text{NH}_4\text{HCO}_3$ was added after milling. The glasses were amorphous and had no significant crystalline phases after milling and the glass particles also exhibited irregular shapes with a wide size distribution (results not shown). The average particle size of S53B50, P40B10 and Sr were 39.9 $\mu$m, 32.8 $\mu$m and 46.3 $\mu$m respectively, with this said, it is noteworthy to mention that the large particle size obtained for these glasses was due to the small force that was used during crushing compared to the force applied during crushing of the glasses that were prepared without $\text{NH}_4(\text{HCO}_3)$. This increase in particle size was crucial in that, fine particles were reported to enhance crystallization of bioactive glass powders.

Densification and Microstructural analysis of samples sintered with/without $\text{NH}_4(\text{HCO}_3)$. Samples were sintered at temperatures between 480 and 600 °C. The obtained densities of the sintered bioactive glasses without $\text{NH}_4(\text{HCO}_3)$ are shown in Fig. 4. The density of the sintered samples for glass S53B50 and Sr was found to increase up to 560 °C and decreased as the sintering temperature was further increased to 600 °C. The decrease in density may be attributed to the inhibition of the viscous flow of the particles caused by surface nucleation and/or crystallization; hence, particles may not fuse together properly during sintering resulting in poor densification of the material.

Glass P40B10 samples showed the same trend, but with slightly lower densification than S53B50 and Sr, this may be due to high activation energy for viscous flow for this particular glass composition. The maximum densification (theoretical density-TD %) achieved for glasses S53B50, P40B10 and Sr were 93%, 72% and 92% respectively.

Figure 5 shows the XRD patterns of the bioactive glasses sintered without $\text{NH}_4(\text{HCO}_3)$ for each sintering temperature. As the sintering temperature was increased, the samples began to crystallize as evidenced by the presence of sharp diffraction peaks. As expected, the crystallization occurs at lower temperature than $T_s$ due to the small particle size used compared to the one analyzed by DTA.

SEM investigations of the sintered bodies' cross sections in Fig. 6 reveal dense microstructures which correlates with the densification results in Fig. 4. However, the presence of micropores is also evident which could have contributed to the reduction in the density of the samples. These micropores might be residual porosity as a result of the incomplete sintering process. In addition, Fig. 7 shows the presence of crystals formed on the glass surface during sintering as a result of crystallization. As mentioned earlier, this action should be controlled/minimized in order to retain the bioactivity of the glasses.

The porosity from the sintered samples without $\text{NH}_4(\text{HCO}_3)$ was too low for tissue engineering application therefore; a second batch of experiments was conducted with the addition of $\text{NH}_4(\text{HCO}_3)$ with the aim of increasing the porosity of the samples. Table 3 summarizes the density and porosity of the obtained scaffolds.
The porosity of the scaffolds was found to increase as the NH₄(HCO₃) content was increased. A similar trend was observed by Locs et al.¹⁶, whereby the open and total porosity were found to increase from 25 to 69% and from 32 to 73% when NH₄(HCO₃) content was varied from 0 to 2.75 wt.%. The glass compositions reached an open porosity of more than 60% at the addition of 70 vol.% NH₄(HCO₃), with glass Sr obtaining the highest open porosity of 72% TD. SEM analysis of the cross sections in Fig. 8 reveal a porous microstructure with well-connected open porosity and irregularly shaped pores; hence, the low density. The formation of such porosity is caused by the high volume expansion (more than 1000 times) ratio during decomposition of NH₄(HCO₃)¹⁶. It is known that porosity decreases the density of the materials as confirmed by the densification results. The interconnection between the pores is clearly presented in the SEM micrographs. Homogeneity of pore distribution is also evident on the cross section of the scaffolds. All the glasses had a similar microstructure.

In addition, the pore walls were composed of dual microstructures (results not shown) with micro pores observed on the pore struts as well as all over the surface; this was probably due to the gas released during the sintering process. However, the micro pores were only clearly visible on glass P40B10 and Sr. The presence of micro

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**Figure 3.** SEM images of (a) S53B50 (b) P40B10 and (c) Sr bioactive glass powder showing the particle size, shape and EDS analysis.
pores is an important feature of these scaffolds, as it is known that small pores favour cell adhesion and allow the physiological fluids to enter the inner part of the scaffold.

The scaffolds were investigated by XRD to check for the presence of any crystalline phase formed during sintering. The XRD patterns for glass S53B50 and P40B10; shown in Fig. 9 had a broad shallow peak which indicates the glasses retained the amorphous state during sintering, while glass Sr still showed a clear sign of crystallization. However it is worth mentioning that the intensity of the diffraction pattern was more pronounced when the glass was sintered without foaming agent.

The particle size is known to play a critical role in sintering as well as in crystallization. Particle sizes for samples prepared without NH₄(HCO₃) were in average 20.5 µm, 18.4 µm and 26.4 µm for glass S53B50, P40B10 and Sr respectively, while the average particle sizes for the glass powders sintered with NH₄(HCO₃) was 39.9 µm, 32.8 µm and 46.3 µm for S53B50, P40B10 and Sr respectively. Fine particles are known to increase the number of nucleation sites by increasing surface to volume ratio thus increasing the crystallization kinetics of glasses with surface initiated crystallization. Aside from the particles size which might affect the crystallization kinetics, the presence of carbon, from the binder, at the particles surface may inhibit crystallization.

For a scaffold to be ideal for any tissue engineering application, the most important architectural feature of the scaffold is the interconnecting pore apertures of the pore network. A modal (Dmode) interconnected pore size of 100 µm or more is required in order for the scaffold to allow tissue ingrowth and vascularization. The 3-D µCT...
images of the structure of the optimized scaffolds at different NH$_4$(HCO$_3$) content are shown in Fig. 10 for glass S53B50. From the images, it is evident that the pores generated and porosity increased with the NH$_4$(HCO$_3$) as observed with the SEM analysis. There is also clear visual evidence of the high interconnectivity and porosity of the scaffolds. In addition, there appears to be a homogenous distribution of pores which are distributed uniformly all over the scaffolds. This trend was observed for glass P40B10 and Sr scaffolds.

Figure 11 presents the pore size distribution of the scaffolds obtained from mercury intrusion porosimetry. The vertical axis is a derivative of the volume of mercury intruded into the scaffolds relative to the pore diameter. The figure shows that all scaffolds have a broad pore distribution in the range 0.003 µm – 100 µm with a D$_{mode}$ at ~50 µm.

Table 4 summarises the characterization data of the scaffolds obtained from the analysis. From the results, as expected, the average pore size and porosity increased as a function of NH$_4$(HCO$_3$) content.

From the results, as expected, the average pore size and porosity increased as a function of NH$_4$(HCO$_3$) content. The pore size obtained from porosimetry is significantly lower than values obtained by µCT. However, one should keep in mind that the porosimetry does not provide measure of the pores but rather, the interconnected size. In addition it is noteworthy to mention the similarities in the porosity obtained in Tables 3 and 4 respectively.

**Discussion**

All the DTA traces exhibited an endothermic event corresponding to the glass transition and an exothermic event corresponding to the crystallization of the glass. From the thermograph, T$_g$ (glass transition temperature) and T$_c$ (onset of crystallization) were recorded and are presented in Fig. 1. $\Delta T$ ($\Delta T = T_c - T_g$), also presented in Table 2, is a gauge to glass resistance to crystallization. According to Brauer, crystallization of bioactive glasses can be controlled by increasing the working range which is the temperature difference between crystallization and the glass transition. This stability range, $\Delta T$, is the temperature range in which glass viscosity allows for sintering,
fiber drawing, or other processing. A large temperature gap between glass transition and onset of crystallization suggests that viscous flow is more likely to occur prior to the crystallization. A small temperature gap between glass transition and onset of crystallization suggests a nucleation close to the glass transition temperature and therefore more risk of nucleation and growth of crystals during sintering, prior or during sintering. Furthermore, it is noteworthy that all glasses are expected to have a surface initiated crystallization as demonstrated for the Sr glass in ref. 22 and for typical silicate bioactive glasses in refs 6 and 8. The surface crystallization of borosilicate glasses was also evident and will be reported shortly 7, 19. It is therefore expected that with small particle size the risk of crystallization upon increasing temperature increases due to an increase in the number of nucleation sites with increasing the surface area to volume ratio6, 8, 24, 25. Glass S53B50 and P40B10 have a wider processing window than glass Sr; therefore, sintering can be achieved without the glasses enduring adverse crystallization during heat treatment.

The XRD patterns presented in Fig. 5 show the transitional behavior which led to the crystallization of the individual glasses. Figure 5(a and b) showed the onset of crystallization at temperatures between 520 °C and 540 °C while Fig. 5(c) showed crystallization transition at 480 °C. With an increase in temperature the diffraction patterns become more intense and distinct, indicating an increase in crystallinity. The SEM micrographs in Fig. 7 display the cross-sectional morphology of the sintered scaffolds, highlighting the uniformity and densification achieved through the sintering process. The table below summarizes the composition, density, theoretical density (TD), and porosity of the sintered scaffolds.

Figure 7. SEM micrographs of the cross section of glass samples sintered at 600 °C for glass (a) P40B10 (b) S53B50 and (c) Sr.

<table>
<thead>
<tr>
<th>Composition</th>
<th>Temperature (°C), Time (Hr)</th>
<th>Density (g/cm³)</th>
<th>Theoretical density (%) TD</th>
<th>Porosity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>S53B50 – 60 vol.%, NH₄(HCO₃)</td>
<td>540, 1</td>
<td>1.07</td>
<td>41</td>
<td>59</td>
</tr>
<tr>
<td>S53B50 – 70 vol.%, NH₄(HCO₃)</td>
<td>540, 1</td>
<td>0.86</td>
<td>33</td>
<td>67</td>
</tr>
<tr>
<td>P40B10 – 60 vol.%, NH₄(HCO₃)</td>
<td>540, 1</td>
<td>1.19</td>
<td>41</td>
<td>59</td>
</tr>
<tr>
<td>P40B10 – 70 vol.%, NH₄(HCO₃)</td>
<td>540, 1</td>
<td>0.95</td>
<td>32</td>
<td>68</td>
</tr>
<tr>
<td>Sr – 60 vol.%, NH₄(HCO₃)</td>
<td>500, 1</td>
<td>1.02</td>
<td>36</td>
<td>64</td>
</tr>
<tr>
<td>Sr – 70 vol.%, NH₄(HCO₃)</td>
<td>500, 1</td>
<td>0.79</td>
<td>28</td>
<td>72</td>
</tr>
</tbody>
</table>

Table 3. Composition, Density and Open porosity of sintered scaffolds.
peaks sharpen and increase in intensity related, as expected, to an increasing crystallinity of the glass. This means that glass S53B50 and P40B10 can be sintered up to 540 °C without undergoing adverse crystallization in contrast to glass Sr, which will undergo crystallization at any sintering temperatures. Furthermore it can also be seen that

Figure 8. SEM micrographs of porous S53B50, P40B10 and Sr bioactive scaffolds mixed with (a), (c) and (e) 60 vol.% (NH₄)HCO₃ and (b, d and f) 70 vol.% (NH₄)HCO₃ respectively, sintered at 540 °C and 500 °C for 1 hour respectively.

Figure 9. XRD patterns of the porous glass scaffolds for S53B50, P40B10 and Sr.
Figure 10. An X-ray micro tomography (µCT) image of S53B50 glass scaffold mixed with (a) 60 vol.% and (b) 70 vol.% NH₄(HCO₃).

Figure 11. Pore size distribution of the glass scaffolds at different NH₄(HCO₃) content.
the density of the samples decreases as the intensity of the peaks increases with temperature. This evidently shows how crystallization effects the densification of amorphous materials.

SEM images of the cross section of the sintered samples revealed a dense microstructure which explains the high theoretical densities obtained between sintering temperatures of 480 °C–560 °C (see Fig. 6). Figure 7 shows the SEM micrographs, recorded with backscattered electrons, of the samples sintered at 600 °C for glass (a) P40B10 (b) S53B50, and (c) Sr at high magnification. There is strong evidence of crystals forming in the P40B10 glass, whereas the darker areas correspond to the crystals and the brighter areas to the remaining glass. This correlates strongly with the corresponding XRD traces, as shown in Fig. 5. SEM investigation of glass S53B50 showed almost no or minor presence of crystals. This is in agreement with the XRD pattern exhibiting low number of diffraction peaks with low intensity indicating reminiscence of a mainly amorphous matrix. The XRD patterns of glass Sr show adverse crystallization and this is clearly confirmed by the SEM micrograph of the glass which reveal high volume of crystals present on the cross section of the samples. Similar results have been reported in ref. 19 where rapid surface crystallization led to fully crystallized particles at such temperatures. In contrast to the crystallization mechanism of glass S53B50, crystals of glass P40B10 appeared to be irregularly shaped and clustered together while crystals of glass Sr exhibit a thatch like shape covering the surface of the sintered body.

One of the most important architectural feature of the scaffold is the pore size. Ideally, the pores should be at least 100 µm to enable cell migration and bone ingrowth26–28. In this work, scaffolds were found to have an average pore diameter in the range of 243.8 – 285.5 µm as reported in Table 4. This pore diameter range is suitable for tissue engineering application as it will not only allow good permeability of the fluids but will also promote bone ingrowth and vascularization. A second architectural feature that is of importance is the size of the interconnections. The interconnections are the pathways between the pores. Porosimetry gives indication of the size of the interconnections. Here all scaffolds have interconnections in the order of 50 µm. It was reported in ref. 29, that the minimal necessary interconnections, in bioceramics, is 20 µm, but it is most favorable for cell penetration when they are >40 µm. Interconnections over 50 µm can assure mineralized bone formation. The similarity in overall porosity obtained from calculation of the theoretical density, analysis of the μCT images and from porosimetry experiments (Tables 3 and 4) also indicates that the majority of the pores are open and interconnected which is favourable for fluid transport throughout the scaffold.

### Methods

#### Glass and Powder preparation.

The starting materials for the borosilicate glass designated S53B50 were analytical reagent grades of Na₂CO₃, H₃BO₃, CaCO₃, CaHPO₄·2H₂O and 99.4% pure SiO₂, while the borophosphate and phosphate glasses designated P40B10 and Sr respectively were prepared using NaPO₃, H₃BO₃, SrCO₃, CaCO₃ and (NH₄)₂HPO₃ as raw materials. Ca (PO₃)₂ and Sr (PO₃)₂ were used as precursors for the phosphate glasses and were prepared independently using a slow heating rate up to 900 °C.

The three glass compositions studied are given in Table 1. Glass S53B50 had a nominal oxide composition of 26.93SiO₂-26.93B₂O₃-22.66Na₂O-1.72P₂O₅-21.77CaO, while glass P40B10 and Sr had a nominal composition of (50-x) P₂O₅-xB₂O₃-20CaO-20SrO-10Na₂O; where x = 0 (Sr) or 10 (P40B10). Proportions of relevant oxides of the starting materials were mixed together in a platinum crucible and melted at 1250 °C for glass S53B50 and 1100 °C for glasses P40B10 and Sr. The melt was poured onto a graphite mould and annealed at 40 °C below their respective glass transition temperature Tg to release internal stresses and allowed to slowly cool down to room temperature in the annealing furnace. The obtained glass ingots were then crushed using a metallic mortar and pestle for 1 hour and sieved.

#### Thermal properties.

The glass transition temperature Tg and the onset crystallization temperature To of the glasses were determined using differential thermal analysis (SDTA, Netzch F1 JUPITER*) at a heating rate of 10 °C/min for particle size of 125–250 µm. The measurements were performed on 30 mg samples in platinum pans in a N₂ atmosphere. The glass transition temperature Tg was taken at the inflection point of the first endotherm, obtained by taking the first derivative of the DTA curve. The crystallization temperature To was found at the maximum of the exothermic peak.
The crushed powders were charged and milled in a planetary ball mill (Fritsch Pulverisette 6) using alumina balls (2 mm diameter) for 1 hour at 150 rpm, with distilled water as a grinding media. The charge to ball ratio was kept at 1:1. Dolapix C64 and PEG (polyethylene glycol) were added at 3 wt.% to the mixture as a dispersant and binder respectively. The slurries were dried in an oven at 70 °C for 24 hours. The dried powders were then sieved and a Mastersizer 2000 (Malvern Instruments, Germany) was used to measure the particle size distribution of the powders.

For the porous scaffolds, the glass powders were mixed with 60 vol.% and 70 vol.% NH₄(HCO₃) foaming agent. To ensure homogeneity, mixing was done in the T2F turbula mixer using 2 mm alumina balls for 30 minutes. After mixing, the resulting powders were compacted using a hydraulic press at 19.6 MPa to obtain pellets of 18.01 mm diameter and 4 mm thick.

**Sintering with/without NH₄(HCO₃).** Sintering of the samples was done with and without NH₄(HCO₃) foaming agent. The samples prepared without NH₄(HCO₃) were sintered using a heating rate of 10 °C/min at sintering temperatures between 480 °C and 600 °C and a holding time of 2 hours. On the other hand the samples with NH₄(HCO₃) were sintered for 1 hour at 540 °C for S53B50 and 500 °C for P40B10 and Sr after burning off the NH₄(HCO₃) foaming agent at 67 °C for 2 hours. All the samples were sintered in a Muffle furnace (Elite Thermal system BRF 18/SE-2416 + 2116) and cooled by switching off the furnace immediately after the sintering time was completed.

**Characterization of sintered samples.** The density of the samples sintered without NH₄(HCO₃) foaming agent was determined using Archimedes principle as most of the pores are closed, whereas the density of the porous scaffolds, obtained with addition of a porogen (NH₄(HCO₃)), was determined by the geometrical density (mass of the sample over its volume). The general equation used to derive the theoretical density of the sintered bodies was:

\[
\text{Theoretical density (\%TD)} = \frac{\text{Calculated density}}{\text{Actual density}} \times 100
\]

The sintered samples were further characterized using X-ray diffraction (D2 Phaser), with Cu Kα radiation. Diffractograms were collected over a 2θ range of 10–90°, with a step size of 0.02°. All the microstructure observations were done using scanning electron microscopy (FEI Quanta 400 FEG SEM) with attached EDS system. Micro-computational tomography (μCT), a MicroXCT-400 (Carl Zeiss X-ray Microscopy, Inc., Pleasanton, USA) was used with tube voltage 140 kV and current 71 μA. Pixel size was 5.6 μm. Porosity analysis was done with Fiji using Bone plug-in. μCT visualizations were done with Avizo 9.1 (FEI Visualization Sciences Group).

Mercury Intrusion Porosimetry (using an AutoPore IV 9500 V1.09, serial 833) and microcomputational tomography (μCT) were performed to determine the porosity and the pore size distribution of the scaffolds. The mercury was intruded using a pressure from 0.10 to 60 000 psi at an equilibrium time of 10 seconds using an evacuation pressure and time of 50 μmHg and 30 minutes, respectively.

**References**


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Author Contributions
All experiments were conducted by E.E., and designed by I.S and J.M. The results and their effects were discussed by E.E., O.J., I.S. and J.M. All authors reviewed the manuscript.

Additional Information
Competing Interests: The authors declare that they have no competing interests.

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